



INTERNATIONAL JOURNAL OF PHARMACEUTICAL RESEARCH AND BIO-SCIENCE

COMPARATIVE ACARICIDAL BIOEFFICACY OF *SPILANTHES ACMELLA* AND *CALOTROPIS PROCERA* OF EASTERN HIMALAYAN REGION AGAINST CATTLE TICK, *RHIPICEPHALUS MICROPLUS*

SUNITA BHASKAR¹, VIVEK SHARMA¹, ANU RAHAL²

1. Department of chemistry, GLA University, Mathura, Uttar Pradesh, 281406, India.

2. Department of animal health division ICAR-CIRG, Makhdoom, Farah, Mathura, Uttar Pradesh, 281122, India.

Accepted Date: 01/10/2019; Published Date: 27/10/2019

Abstract: In this study, relative toxicity of *Spilanthes acmella* and *Calotropis procera* was evaluated against adults and larvae of *Rhipicephalus (Boophilus) microplus*. The aerial part of both plants materials were collected from Eastern Himalayan Region (West Bengal) of India. Plant materials were washed, shade dried, coarsely ground, methanol extracted and dried by rotary evaporator and collected proper yield of extracts. The crude methanolic extracts were further fractionated using solvents (hexane, ethyl acetate, chloroform) of different polarity and finally aqueous fraction was collected and dried. Methanolic crude extracts and their fractions (hexane, ethyl acetate, chloroform and aqueous) concentrations of both the plants were tested against the engorged adult females and cultured larvae of *Rhipicephalus (Boophilus) microplus*. The bioefficacy observations are shown in table 3 and mentioned LC₅₀, LC₉₀ and their related statistics. Adult and larval stages were significantly affected by the chloroform extract of both the plants selected and observed the most potent with LC₅₀ 50.22 and 13.86 mg/ml of *Calotropis procera* and LC₅₀ 60.94 and 25.82 mg/ml of *Spilanthes acmella*.

Keywords: *Spilanthes acmella*, *Calotropis procera*, *Rhipicephalus (Boophilus) microplus*, Acaricidal bioefficacy.

Corresponding Author: SUNITA BHASKAR



PAPER-QR CODE

Access Online On:

www.ijprbs.com

How to Cite This Article:

Sunita Bhaskar, IJPRBS, 2019; Volume 8(5): 39-49

INTRODUCTION

Ticks are the main problem of the livestock industry in tropical and subtropical regions of the world and they are considered as vectors of many infectious diseases to animals and man. It has been reported that approximately 75% of livestock animals are at risk for ticks and tick-borne diseases causing a global annual loss of 7000 million American dollars [1, 2, 3]. The severe tick infestation may lead to anaemia, reduction in weight gain, direct damage to cattle skins and related production losses in addition to transfer of various tick borne diseases in animals and human beings viz. Lyme disease, Rocky mountain, Spotted fever, Relapsing fever, Tularemia, Meningoencephalitis, Colorado tick fever, Crimean-congo hemorrhagic fever, Bovine Babesiosis, Anaplasmosis, Cytauxzoonosis, etc. [4, 5].

Control of ticks is usually based on chemical acaricidal applications including synthetic pyrethroids, organophosphates, amitraz, pyrethroids and macrocyclic lactones and Ivermectin, Moxidectin, Doramectin and Abamectin etc. Most of these acaricides possess harmful effects on animal health and environment and their uses have caused great concern in society and government [6, 7]. Due to indiscriminate use of these conventional pesticides, number of incidences of resistance have been reported worldwide [8, 9, 10, 11, 12, 13, 14]. Now a days, tick control has become a challenge to researchers around the world, who seek a sustainable way to do it. Synthetic acaricides have been used widely and extensively, however, the high cost, the hazardous effects on environment, mainly human beings are the indirect target [15].

The time is, therefore, demanding to develop sustainable, ecofriendly and cost effective alternative, especially for the marginal farmers with limited capital who constitutes the majority of animal rearers in developing countries. The use of phytoextracts is a sustainable option to synthetics as they are biodegradable, cost effective and efficient. Several plant extracts with acaricidal effects have reported worldwide [16, 17, 18, 19]. Among the natural products, plant extracts and essential oils have shown significant activity against all the stages of economically important tick species resistible as compared to synthetic pesticides [20, 21, 22, 23, 24, 25, 26]. Several Indian plants have been mentioned in literature to show promising efficacy against ticks. The present study is, therefore, intended to evaluate the acaricidal properties of two traditional used medicinal plants, *Calotropis procera* and *Spilanthes acmella* against *Rhipicephalus microplus*.

MATERIALSAND METHODS

Collection extraction of plant materials

Fresh aerial parts of *Calotropis procera* and *Spilanthes acmella* (Figure 1) were collected from Eastern Himalayan region of India and washed, shade dried at room temperature and grinded

by stainless steel grinder as coarse powder. The coarse powders were extracted in soxhlet assembly in methanol solvent and dried in rotary evaporator to get proper yield. *Calotropis procera* and *Spilanthes acmella* plants methanolic crude extract were separated in 3 different solvents and distilled water. 10 g methanolic crude extract was dissolved in 40 ml distilled water into 100 ml glass beaker. The solution was transferred into separating flask and added 60 ml hexane, mixed well and left until 30 to 60 minutes for separation of hexane layer. It was collected in 100 ml glass bottle (upper layer was hexane) and rest amount of 40 ml aqueous part transferred into separating flask and added 60 ml ethyl acetate, mixed it and left 30 to 60 minutes untouched, ethyl acetate layer collected in 100 ml glass bottle (upper layer was ethyl acetate) and rest amount of 40 ml aqueous part transferred into separating flask and added 60 ml chloroform, mixed it and left 30 to 60 minutes undisturbed for separation of two layers. After that, chloroform layer and aqueous part collected in 100 ml two glass bottles (lower layer was chloroform). Hexane, ethyl acetate and chloroform layers dried at room temperature to remove the solvents and distilled water and to get the fractions.

Bioassay

The dropped ticks after taking full blood meal from the animal's body during night were collected early in the morning. Ticks were collected from farmer's house from Salempur and Makhdoom village in plastic containers and covered with cotton cloth for proper oxygen supply. Ticks were washed in running tap water to remove dust and other impurity on the body surface and dried by filter paper after then keep in refrigerator (4°C) for further study. In adult immersion test (AIT), 105 ticks were divided into seven groups, six groups treated one kept as control and 15 ticks in each group having weight approximately 2 to 3g. Ticks were taken in petri dishes of 9 cm in diameter containing bed of filter paper (Whatman No.1) and treated group immersed into the respective test concentrations of methanolic crude extract and fractionations (hexane, ethyl acetate, chloroform and aqueous parts) and the control group immersed in distilled water and time of test approximately 2-3 minutes. Treated ticks placed on petri dishes and incubated into BOD incubator at $28 \pm 2.0^\circ\text{C}$ temperature and 70-80 % relative humidity, mortality data observed after 24 hours and died ticks removed from petri dishes and observed in the same way up to 15 days until complete oviposition [27]. 5 days old larvae were used [28] in larval immersion test (LIT) test and larvae were divided to seven groups (n=100-150). Larvae were picked with a help of paint brush No. 4 and gently transferred to clean 1 ml tubes covered with muslin cotton clothes and tied with rubber band. Each group was immersed into the methanolic crude extract and fractionations concentrations while the control group was immersed in distilled water and test time for 2-3 minutes. The treated tubes were placed in 100 ml glass beaker and incubated for 24 hours in BOD at $28 \pm 2.0^\circ\text{C}$ temperature and 70-80% relative humidity. Mortality data was observed after 24 hours [27]. LC_{50} and LC_{90} values were

calculated by using to Probit Analysis [29] and other statistical parameters by using software developed by Indian Institute of Chemical Technology, Hyderabad [30].



Figure 1: *Spilanthes acmella* and *Calotropis procera*

RESULTS AND DISCUSSION

The bioefficacy observations of methanolic crude extracts and its fractions of *Calotropis procera* and *Spilanthes acmella* are shown in table 1 and mentioned LC_{50} , LC_{90} and their related statistics. Chloroform extract of *Calotropis procera* was found the most effective against both adult and larval stages with LC_{50} 50.22 mg/ml and LC_{90} 156.48 mg/ml (adult) and LC_{50} 13.86 mg/ml and LC_{90} 47.68 mg/ml (larvae). In case of *Spilanthes acmella*, chloroform fraction was again observed the most potent against both adults and larvae with LC_{50} 60.94 mg/ml and LC_{90} 195.08 mg/ml (adult) and LC_{50} 25.82 mg/ml and LC_{90} 74.50 mg/ml (larvae). In present study methanolic crude extracts and their fractions hexane, ethyl acetate, chloroform and aqueous concentrations both of plants were tested against the engorged adult females and cultured larvae of *Rhipicephalus (Boophilus) microplus* and chloroform fractions of both the plants selected were observed to be the most effective.

Table 1: Relative acaricidal toxicity of *Calotropis procera* and *Spilanthes acmella* against different stages of *Rhipicephalus (Boophilus) microplus* after 24 hours of treatment.

Plants name	Target stage	Extracts	χ^2	Regression equation Y=Intercept X + Reg. Coeff.	$LC_{50} \pm SE$ (UFL-LFL) (mg/ml)	RT	$LC_{90} \pm SE$ (UFL-LFL) (mg/ml)	RT
<i>Calotropis procera</i>	Adult	Methanol crude	5.70	Y=0.99X+1.93	129.82±21.53 (172.03-87.62)	2.59	600.71±293.92 (1176.79-24.62)	3.84
		Hexane fraction	4.75	Y=5.25X+3.44	95.39±10.77 (116.50-74.28)	1.89	224.89±35.55 (294.56-	1.44

							155.21)	
		Ethyl acetate fraction	2.37	$Y=3.77X+3.14$	61.78±9.59 (80.57-42.99)	1.23	157.96±24.26 (205.51-110.40)	1.01
		Chloroform fraction	3.04	$Y=2.01X+2.59$	50.22±10.69 (71.64-29.27)	1	156.48±28.60 (212.53-100.42)	1
	Larvae	Aqueous fraction	1.22	$Y=0.89X+1.97$	96.99±17.04 (130.39-63.59)	1.93	432.34±170.77 (767.04-197.64)	2.76
		Methanol crude	12.16	$Y=0.98X+1.61$	30.79±1.85 (34.43-27.16)	2.22	191.57±18.17 (227.19-155.95)	4.02
		Hexane fraction	38.74	$Y=0.94X+2.29$	38.99±1.47 (41.88-36.11)	2.81	141.28±8.04 (157.04-125.52)	2.96
		Ethyl acetate fraction	39.59	$Y=0.95X+1.83$	16.50±1.44 (19.33-13.67)	1.19	83.10±4.55 (92.01-74.19)	1.74
		Chloroform fraction	25.09	$Y=0.12X+2.39$	13.86±1.11 (16.03-11.69)	1	47.68±1.90 (51.41-43.95)	1
		Aqueous fraction	17.07	$Y=0.71X+2.12$	49.33±1.75 (52.75-45.90)	3.56	198.22±15.09 (227.81-168.64)	4.16
		<i>Spilanthes acmella</i>	Adult	Methanol crude	9.58	$Y=5.99X+3.21$	259.67±29.91 (318.29-201.04)	4.26
	Hexane fraction		2.07	$Y=1.39X+2.27$	65.16±10.96 (86.65-43.68)	1.07	238.63±56.78 (349.92-127.35)	1.22
	Ethyl acetate fraction		2.15	$Y=2.49X+2.65$	66.83±11.38 (89.15-44.52)	1.09	203.11±39.46 (280.45-125.77)	1.04

	Chloroform fraction	2.30	$Y=2.06X+2.54$	60.94±11.44 (83.37-38.52)	1	195.08±38.69 (270.93-119.23)	1
	Aqueous fraction	4.27	$Y=1.52X+2.31$	65.27±12.76 (90.28-40.25)	1.07	233.19±54.52 (340.05-126.34)	1.19
Larvae	Methanol crude	70.82	$Y=0.74X+2.34$	28.61±1.32 (31.20-26.01)	1.39	101.12±4.76 (110.44-91.79)	1.36
	Hexane fraction	101.98	$Y=1.24X+2.57$	26.46±1.12 (28.64-24.27)	1.29	83.23±3.12 (89.35-77.12)	1.12
	Ethyl acetate fraction	65.69	$Y=0.08X+2.19$	20.56±1.26 (23.02-18.09)	1	78.74±3.49 (85.57-71.91)	1.06
	Chloroform fraction	48.58	$Y=1.72X+2.78$	25.82±1.04 (27.85-23.78)	1.26	74.50±2.58 (79.56-69.45)	1
	Aqueous fraction	24.04	$Y=0.81X+1.63$	37.11±1.96 (40.96-33.27)	1.80	226.93±23.86 (273.69-180.16)	3.05

RT: Relative toxicity

Acaricidal properties of different botanical derivative have been reported against the different species of the ticks by different researcher from different parts of the world. The acaricidal effect of crude ethanolic extract of *Acmella oleracea* (L.) R.K. Jansen aerials parts at different concentrations on *Amblyomma cajennense* (Fabricius, 1787) (Acari: Ixodidae) ticks was reported by [31, 32] studied the acaricidal effect of *Synadenium glaucescens* (Euphorbiaceae) extracts on *Boophilus decoloratus* and *Boophilus microplus*. *In Vitro* studies on *Rhipicephalus sanguineus* in Abomey-Calavi has been reported the effect of ethanolic extract of *Tephrosia Vogellii* [33]. The relative toxicity against adult ticks and larvae of *Rhipicephalus (Boophilus) microplus* has been reported for methanolic extract of *Adhatoda vasica* roots by [34]. [35] reported anti-tick activity against *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae) and identification of potential plant extracts. Observations on the crude extracts of *Cymbopogon wintericum* Jowett and *Azadirachta Indica* efficacy against *Rhipicephalus (Boophilus) microplus* and inhibited of oviposition 94.29 % mortality were made by [36]. Acaricidal property of hexane extract of *Adhatoda vasica* roots against *Rhipicephalus (Boophilus) microplus*, has been reported by [37]. The extract of *Carapa guianensis* seed oil, leaf essential oil of *Cymbopogon*

martini and crude extract of *piper tuberculatum* [38]. [39] were reported 10% (100000 ppm) and 20% (200000 ppm) as effective concentrations of leaf and stem for AIT and oviposition inhibition, respectively of methanol extract of *Petiveria alliacea* against *Rhipicephalus (Boophilus) microplus*.

The results of the present work showing the promising acaricidal activities against the target organism and might be used as an potential alternative to control the ticks. The efficacy of *Calotropis procera* and *Spilanthes acmella* methanolic crude extracts and its fractionation have been observed against *Rhipicephalus (Boophilus) microplus*. The target organism revealed a dose dependent pattern as expected and the study indicates that the plants *Calotropis procera* and *Spilanthes acmella* have a great potential as an efficacious bioacaricide and needs to be explored for avoiding indiscriminate pesticide use in animals. So, further study should be carried out in future to isolate the specific chemicals constituents and pharmacological activity will be carried out in proper scientific way.

ACKNOWLEDGEMENT

The author is thankful to the Director and Head, Department of Chemistry, GLA University, Mathura, and the Director and Head, Animal Health Division, Central Institute for Research on Goats, Makhdoom, Farah, Mathura for the guidance and the cooperation and the UGC-RGNF fellowship financial support.

REFERENCES

1. Van den Broek, A.H., Huntley, J.F., Halliwell, R.E., Machell, J., Taylor, M. & Miller, H.R. (2003). Cutaneous hypersensitivity reactions to *Psoroptes ovis* and Der p 1 in sheep previously infested with *P.ovis* - the sheep scab mite. *Veterinary Immunology and Immunopathology* 91:105-117.
2. Wall, R. (2007). Ectoparasites: Future challenges in a changing world. *Veterinary Parasitology* 148: 62-74.
3. Ghosh, S., Bansal, G.C., Gupta, S.C., Ray, D., Khan, M.Q., Irshad, H., Shahiduzzaman, M., Seitzer, U. & Ahmed, J.S. (2007). Status of tick distribution in Bangladesh, India and Pakistan. *Parasitology Research* 101:S207-S216.
4. Ghosh S., Azhahianambi P. and de la Fuente J. (2006). Control of ticks of ruminants with special emphasis on livestock farming system in India: present and future possibilities for integrated control—a review. *Exp Appl Acarol.* 40:49–66.
5. Ghosh S. and Nagar G. (2014). Problem of ticks and tick-borne diseases in India with special emphasis on progress in tick control research: a review. *J Vector Borne Dis.* 51:259–270.

6. Mansour, F., Azaizeh, H., Saad, B., Tadmor, Y., Abo, M.F. & Said, O. (2004). The potential of Middle Eastern flora as a source of new safe bio-acaricides to control *Tetranychus cinnabarinus*, the carmine spider mite. *Phytoparasitica* 32: 66-72.
7. Chagas, A.C.S., Leite, R.C., Furlong, J., Prates, H.T. & Passos, W.M. (2003). Sensibilidade do carrapa to *Boophilus microplus* solvents. *Ciencia Rural* 33: 109-114.
8. Kumar S., Paul S., Sharma A.K., Kumar R., Tewari S.S., Chaudhuri P., Ray D.D., Rawat A.K.S. and Ghosh S. (2011). Diazinon resistant status in *Rhipicephalus (Boophilus) microplus* collected from different Agro-climatic zones of India. *Veterinary Parasitology* 181: 274-281.
9. Kumar S., Sharma A.K., Ray D.D. and Ghosh S. (2014). Determination of discriminating dose and evaluation 4 of Amitraz resistance status in different field isolates 5 of *Rhipicephalus (Boophilus) microplus* in India. *Experimental and Applied Acarology* 63: 413-422.
10. Sharma A.K., Kumar R., Kumar S., Nagar G., Singh N.K., Rawat S.S., Dhakadd M.L., Rawat A.K.S., Ray D.D. and Ghosh S. (2012). Deltamethirn and cypermethrin resistance status of *Rhipicephalus (Boophilus) microplus* collected from six Argo-climatic regions of India. *Veterinary Parasitology* 188: 337-345.
11. Shyma K.P., Kumar S., Sharma A.K., Ray D.D. and Ghosh S. (2012). Acaricide resistance status in Indian isolates of *Hyalomma anatolicum*. *Experimental and Applied Acarology* 58: 471-481.
12. Kumar R., Nagar G., Paul S., Sharma A.K., Kumar S., Tewari S.S., Ray D.D., Chaudhuri P. and Ghosh S. (2013). Survey of pyrethroids resistance in Indian isolates of *Rhipicephalus (Boophilus) microplus*: identification of C190A mutation in the domain II of the para-sodium channel gene. *Acta Tropica* 125: 237-245.
13. Singh N.K., Jyoti, Haque, Rath M. and Ghosh S.S. (2010). Studies on acaricide resistance in *Rhipicephalus (Boophilus) microplus* against synthetic pyrethroids by adult immersion test with a discriminating dose. *Journal of Veterinary Parasitology* 24: 207-208.
14. Singh N.K., Jyoti, Haque, Singh M., Rath H. and Ghosh S.S. (2014). A comparative study on cypermethirn resistance in *Rhipicephalus (Boophilus) microplus* and *Hyalomma anatolicum* from Punjab (India). *Ticks and Tick Borne Diseases* 5: 90-94.
15. Furlong, J. (1993). Controle do carrapato dos bovinos na Região Sudeste do Brasil. Cadastro Técnico da Escola de Veterinária da Universidade Federal de Minas Gerais 8: 49-61.

16. Chagas, A.C.S., Passos, W.M., Prates, H.T., Leite, R.C., Furlong, J. & Fortes, I.C.P. (2002). Acaricide effect of *Eucalyptus* species essential oils and concentrated emulsion on *Boophilus microplus*. *Brazilian Journal of Veterinary Research and Animal Science* 39: 247-253.
17. Fernandes, F.F., Freitas, E.P.S., Costa, A.C. & Silva, I.G. (2005). Larvicidal potential of *Sapindus saponaria* to control the cattle tick *Boophilus microplus*. *Pesquisa Agropecuária Brasileira* 40: 1243-1245.
18. Fernandes, F.F. & Freitas, E.P.S. (2007). Acaricidal activity of an oleoresinous extract from *Copaifera reticulata* (Leguminosae: Caesalpinioideae) against larvae of the southern cattle tick, *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). *Veterinary Parasitology* 147: 150-154.
19. Zahir, A.A., Rahuman, A.A., Bagavan, A., Santhoshkumar, T., Mohamed, R.R., Kamaraj, C., Rajakumar, G., Elango, G., Jayaseelan, C. & Marimuthu, S. (2010). Evaluation of botanical extracts against *Haemaphysalis bispinosa* Neumann and *Hippobosca maculata* Leach. *Parasitology Research* 107: 585-592.
20. Iori, A., Grazioli, D., Gentile, E., Marano, G., Salvatore, G. (2005). Acaricidal properties of the essential oil of *Melaleuca alternifolia* Cheel (tea tree oil) against nymphs of *Ixodes ricinus*. *Vet. Parasitol.* 129: 173–176.
21. Ribeiro, V.L., dos Santos, J.C., Bordignon, S.A., Apel, M.A., Henriques, A.T., von Poser, G.L. (2010). Acaricidal properties of the essential oil from *Hesperozygis ringens* (Lamiaceae) on the cattle tick *Rhipicephalus (Boophilus) microplus*. *Bioresour. Technol.* 101: 2506–2509.
22. Ribeiro, V.L.S., dos Santos, J.C., Martins, J.R., Schripsema, J., Siqueira, I.R., von Poser, G.L., Apel, M.F A. (2011). Acaricidal properties of the essential oil and precocene II obtained from *Calea serrata* (Asteraceae) on the cattle tick *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). *Vet. Parasitol.* 179: 195–198.
23. Rosado-Aguilar, J.A., Aguilar-Caballero, A., Rodriguez-Vivas, R.I., Borges-Argaez, R., Garcia-Vazquez, Z., Mendez-Gonzalez, M. (2010). Acaricidal activity of extracts from *Petiveria alliacea* (Phytolaccaceae) against the cattle tick *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). *Vet. Parasitol.* 168: 299–303.
24. Ghosh, S., Sharma, A.K., Kumar, S., Tiwari, S.S., Rastogi, S., Srivastava, S., Singh, M., Kumar, R., Paul, S., Ray, D.D., Chaudhuri, P., Rawat, A.K.S. (2011). *In vitro* and *in vivo* efficacy of *Acorus calamus* extract against *Rhipicephalus (Boophilus) microplus*. *Parasitol. Res.* 108: 361–370.
25. Zaman M.A., Iqbal Z., Abbas R.Z., Khan M.N., Muhammad G., Younus M. and Ahmed S. (2012). *In-vitro* and *in-vivo* acaricidal activity of a herbal extract. *Vet. Parasitol.* 186: 431-436.

26. Chungsamamyart N., Jiwajinda S., Ratanakreetakul C. and Jasawan W. (1991). Practical extraction of sugar apple seeds against tropical cattle tick. *Kasetsart J. (Nat. Sci.)*:25: 101-105.
27. Elango G., Rahuman A.A., Kamaraj C., Bagavan A. and Zahir A.A. (2011). Screening for feeding deterrent activity of herbal extracts against the larvae of malaria vector *Anopheles subpictus* Grassi. *Parasitol Res.* In press.
28. Drummond R.O., Ernst S.E., Trevino J.L., Gladney W.J. and Graham O.H. (1973). *Boophilus annulatus* and *Boophilus microplus*: laboratory tests for insecticides. *J Econ Entomol.* 66:130–133.
29. Finney D.J. (1971). *Probit Analysis* (Cambridge University Press, Cambridge). 333.
30. Reddy P.J., Krishna D., Murthy U.S. and Jamil K. (1992). A microcomputer FORTRAN program for rapid determination of lethal concentration of biocides in mosquito control. *CABIOS* 8: 209-213.
31. Anholeto L. A., Oliveira P. R. de., Rodrigues R. A. F., Spindola C. dos S., Labruna M. B., Pizanod M. A., Castro K. N. de C. and Camargo-Mathiasa M. I. (2016). Potential action of extract of *Acmellaoleracea* (L.) R.K. Jansen to control *Amblyomma cajennense* (Fabricius, 1787) (Acari: Ixodidae) ticks. *Ticks and Tick-borne Diseases* 1-8.
32. Nyigo V. A., Mdegela R.H., Malebo H. M., Mabiki F.P. and Fouche G. (2016). Evaluation of acaricidal efficacy of *Synadenium glaucescens* (Euphorbiaceae) against *boophilus* species. *Journal of Medicinal Plants Research* 10, 21:278-285.
33. Jacques D. T., Safiou A., Jédirfort H. and Souaïbou F. (2015). *In vitro* effect of the ethanolic extract of *Tephrosia Vogellii* on *Rhipicephalus Sanguineus* in Abomey-Calavi. *Avicenna Journal of Phytomedicine* 5, 3: 247-259.
34. Kishore V., Srivastava C.N., Kumar A. and Mohan L. (2015). Relative acaricidal bioefficacy of certain root extracts of *Adhatoda Vasica* against *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). *International journal of pharmaceutical research and bio-science* 4, 4: 224-235.
35. Ghosh S., Tiwari S.S., Kumar B., Srivastava S., Sharma A.K., Kumar S., Bandyopadhyay A., Julliet S., Kumar R. and Rawat A.K.S. (2015). Identification of potential plant extracts for anti-tick activity against acaricide resistant cattle ticks, *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). 66:159–171.
36. Peixoto E.C.T.M., Figueiredo A., Novo S.M.F., Porto E.P., Valadares F., Silva L.P. and Silva R.M.G. (2013). Application of *Cymbopogon winterianus* jowitt and *Azadirachta indica* A. Juss in control of *Rhipicephalus (Boophilus) microplus*. *Academic Journals* 7, 32:2392-2398.

37. Madhumitha G., Rajkumar G., Roopan S.M., Rahuman A.A. and Priya K.M. (2012). Acaricidal, Insecticidal activity of fruit peel aqueous extract of *Annonasquamosa* and its compounds against blood feeding parasites. *Parasitology research* 111: 2189-2199.
38. Chagas A.C., de Souza., de Barros L.D., Continguiba F., Furlan M., Giglioti R., Oliveira M.C., de Sena and Bizzo H.R. (2012). *In-vitro* efficacy of plant extracts and synthesized substances on *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). *Parasitology research* 110, 1: 295-303.
39. Rosado-Aguilar J.A., Aguilar-Caballero A., Rodriguez-Vivas R.I., Borges-Argaez R., Garcia-Vazquez Z. and Mendez-Gonzalez M. (2010). Acaricidal activity of extracts from *Petiveria alliacea* (Phytolaccaceae) against the cattle tick, *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). *Veterinary Parasitology* 168: 299-303.